



## Detection of ESBL *E.coli* That Carried STX1 and STX2 Form Common Carp (*Cyprinus carpio*) in Salhaldeen Province

Qusai Saleh Jumma

Department Pathology and Poultry, Faculty of Veterinary Science, Tikrit University, Iraq.

*E.coli* is the most important bacteria that contaminates of fish farms and leads to Pollution and corruption of fish, which causes a threat to public health, the current study aimed to find the distribution of ESBL *E.coli* that carried STX1 and STX2 from common carp (*Cyprinus carpio*) in Salhaldeen province, for this purpose 100 sample were collected from fish, traditional and genetic methods were used. The results of the current study reveal to that Out of 100 fish sample, *E. coli* isolated from 48 in rate of 48% , and 19 isolates out of 48 were diagnosed as ESBL *E.coli* in rate of 39.5% , according to PCR test Stx1 gene detection on 31 isolates out of 48 isolates in rate of 64.5% while Stx2 gene detection on 39 isolates in the rate of 81.2%. We can conclude the high contamination rate of fish and its farms with *E.coli* in Salhaldeen province, most isolates are ESBL, Stx2 gen is more frequent than Stx1 gene.

**Keywords:** *E. coli*, *Cyprinus carpio*, ESBL, shigatoxi

### Introduction

Because of the different diets and health cultures, people's consumption of fish has increased, This is because it has a large amount of vitamins, minerals, and fatty acids, fish is considered as source of about 30% of animal proteins, Despite all these benefits, it can be a carrier of many pathogens that affect humans [1]. *E.coli* is considered as one of the most important bacterial contaminants that may be considered as an indicator of contamination and spoilage of meat [2, 3, 4]. *E.coli* are Gram-negative bacteria, spherical to ciliary-shaped with rounded ends, non-spores forming, move by peritrichous flagella, arranged singly or in pairs. facultative aerobic or anaerobic, possesses both fermentative and oxidative pathways of metabolism, the optimum temperature for its growth is 37 C. It many carbohydrates such as lactose, glucose, mannitol, maltose, and arabinose, producing acid and gas, oxidase test , citrate consumption test, hydrogen sulfide production, and it can analyze urea, , and it is positive for catalase, methyl red, and nitrate

reduction tests [5]. Due the use of antibiotics, several antibiotics resistant bacteria strains were isolated, and the most important of these antibiotics are the beta-lactam group. Bacteria have several mechanisms for antibiotic resistance, these bacterial strains and their resistance mechanism may be transmission to humans by taking of contamination food and water or due to environmental pollution with these bacteria [6, 7, 8]. Shiga toxin one of most important virulence factors of *E.coli*, it takes his name due to its similarity with Shigella toxin except one amino acid [9, 10]. *E.coli* producing Shiga toxin is one of most important causes of enteric disease, shiga tosin play important role in pathogenicity of *E.coli* that causes food poisoning [11, 12].

### Material and Methods

**Samples:** 100 swabs from fish skin gills and intestine were collected from the local market in Salhaldeen province.

\*Corresponding author: Qusai S. Jumma1, E-mail: [qusaisaleh@tu.edu.iq](mailto:qusaisaleh@tu.edu.iq) Tel.: +964 772 504 8881

(Received 15/12/2023, accepted 08/01/2024)

DOI: 10.21608/EJVS.2024.255648.1728

©2024 National Information and Documentation Center (NIDOC)

Swabs cultured on trypton soya broth (Himedia-India) and cultivated at 37C for 24h then loop full subcultured on macConkey agar (Himedia- India) and cultivated at 37C for 24h. suspected colony were sub cultured on Eosin methylene blue (Himedia-India) and cultivated at 37C for 24h, then Gram stain and group of biochemical tests were done according to [13].

**Detection of ESBL *E.coli*** : single purified *E.coli* coloy cultured on chromagar (CHROMagar™ ESBL- Chromogenic media Pioneer – UK).

Detection of shiga toxin producing done by following strategies:

**a. DNA extraction:** were done by use of (DNA Preparation Kit PP-206-Jena Biosciences, Jena, German) and according to Manufacturer's instructions.

**b.Primer:stx1:** F:TCTTGCGGTACTCTAGTAG

R:AGAACGCCCACTGAGATCATC (give out product in size of 180bp).

**c.Stx2:** F: AGAACGCCCACTGAGATCATC, R: TCGCCAGTTATCTGACATTCTG (give out product in size of 255bp). [14].

### **Results and Discussions**

Out of 100 fish samples, *E.coli* was isolated from 48 at rate of 48%, all isolate appeared as gram negative bacteria, fermented to lactose sugar which appear as pink colony on MacConkey agar(Fig. 1) while the colony appear as give green metallic

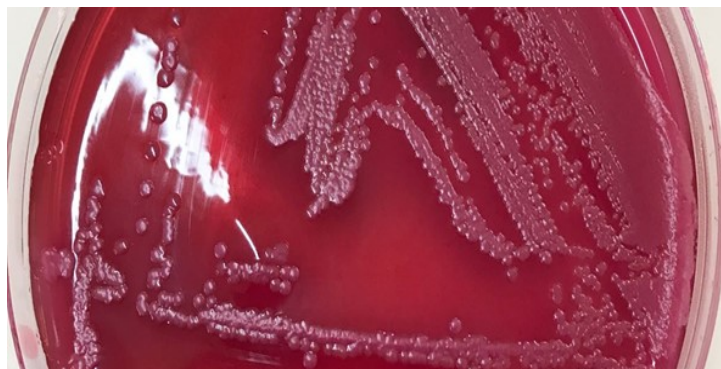
colony on EMB agar figure (2). All *E.coli* isolates give positive to catalase, MR test and Indol test while negative results on the oxidase test, VP, and citrate test.

The isolation rate is less than the results recorded by Alttai *et al.* [15] in the local market in Mousal , Nineveh governorate, Iraq, In the study of Taha and Yassin's [16] isolated *E.coli* in rate of 39% from *Cyprinus carpio* fish in Dohuk province, Iraq. The isolation of *E.coli* from fish refers to water pollution from sources such as Wastewater, waste, or the sack of feed provided to fish contaminated with bacteria [17].

When *E.coli* re cultured on CHROMagar™ ESBL, ESBL *E.coli* diagnosed in 39.5% (19 out of 48) as in figure (3)

In the study of Mahmmoud and Al-Dabbagh [18] shows that 34% of *E. coli* isolated from fish in Mosul are resistant to cefotaxime, while in the study of Kumar *et al.* [19] showed that 38% of fish were contaminated with ESBLs *E. coli*. While in the study of Tyasningsih *et al.* [20] showed that ESBLs *E. coli* formed 1.7% of all *E. coli* isolated from animal products in Indonesia.

According to the PCR test results , Stx1 gene detection on 31 isolates out of 48 isolates at the rate of 64.5% (Fig. 4) while Stx2 gene detection on 39 isolates at rate of 81.2% (Fig. 5).



**Fig. 1. MacConkey agar , shows pink colony of *E.coli***



Fig. 2. EMB agar , shows green metallic sheen colony of *E.coli*

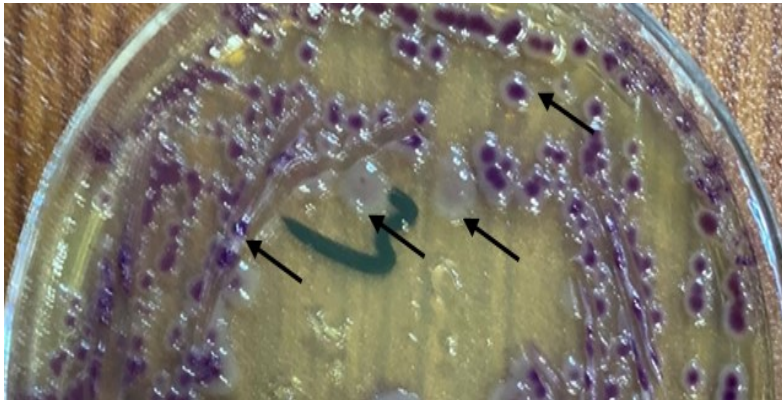


Fig. 3. CHROMagar™ ESBLE agar, shows purple colony of ESBL *E.coli*

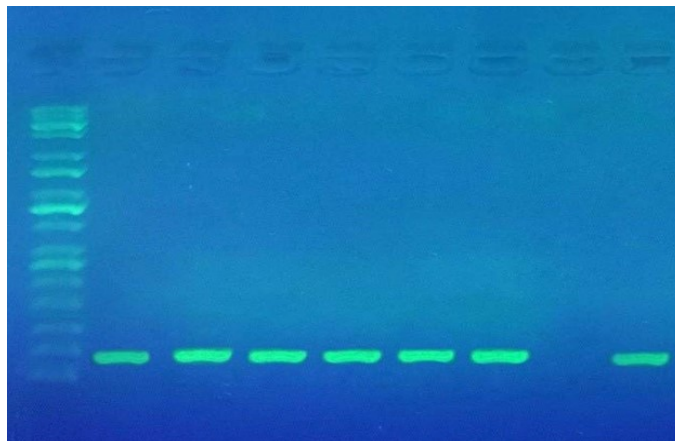
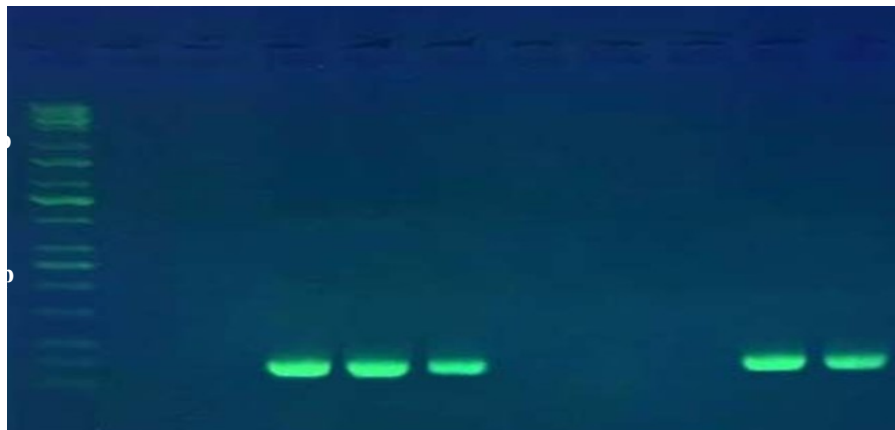


Fig. 4. Results of *E.coli* producing *stx1* gene, Lens 1,2,3,4,5,6, and 8 positive results with band in size of 180bp . M:100pb DNA marker.



**Fig. 5. Results of *E.coli* producing stx2 gene, lens 3,4,5, 9 and 10 positive results with band in size of 255bp, M:100pb DNA marker.**

In study of MS *et al.* [21] showed that *E.coli* produced stx1 gene and stx2 gene in rates of 60% and 70% when isolated from Fish Markets in Egypt, while in study of Pradhan *et al.* [22] showed that the presence of *E.coli* isolates producing stx1 gene and stx2 gene in rate of 44% and 77%, these difference may be due geographic differences of study location. stx-1 and stx-2 seem to be important in human infections. *E. coli* O157:H7 is the principal serotype of this group Enteroinvasive *E. coli* (EIEC) causes a diarrheal illness similar to shigellosis [23,24]. This result may be attributed to the possible attachment and penetration of *E. coli* O157 from 209 mucus into the skin. Although the mucopolysaccharide structure of the mucus layer in the fish 210 skin is reported to protect fish against infections [25] indicated that *E. coli* O157: H7 cells were able to penetrate through the mucus layer of 212 fish and become a source of contamination during processing. [26] Howbeit, injury of the fish 213 body during stressful harvesting and direct contact with the contaminated ice during storage 214 could lead to fish muscle contamination.

### **Conclusion**

We can conclude the high contamination rate of fish and its farms with *E.coli* in Salhaldeen province. most isolates are ESBL, Stx2 gene are more frequent than Stx1 gene.

### **Acknowledgments**

The authors thanks to vet. med. college, Tikrit University. the authors are very grateful to Dr. Bashir S. Nomi and Dr. Ali A. Abd for a limited support throughout the experiment.

### **Conflict of interest**

There are no conflicts of interest to be declared.

### **Funding statement**

The article was not financially supported.

### **Author contributions**

Conceptualization, study design, sample collection, data analyses, Manuscript drafting, and manuscript finalization: Qusai Saleh Jumma

### **References**

1. Abraha, B.H., Admassu, A., Mahmud, N., Tsighe, X.W., Shui, A. and Fang, Y. Effect of processing methods on nutritional and physicochemical composition of fish: a review, *MOJ Food Processing and Technology*, **6**(4), 376–382(2018).
2. Ahmed, N.A., Mahmood, S.S. and Abbas, A.H. A comparative study of some virulence factors and phylogenetic characterization of *Escherichia coli* isolates causing urinary tract infection and the commensal gut. *The Iraqi Journal of Agricultural Science*, **50**(3), 1193-1198(2019).
3. Ayat, A.A. and Shakir, K.A. Functional properties of catfish skin collagen hydrolysates. *Iraqi Journal of Agricultural Sciences*, **52**(6), 1528-1540 (2021).
4. Fatemeh Andalib, Hojjat Baghshahi, Memarzadeh, Mohammadreza and Hosein, Akbari. In vitro and In vivo Effects of Zataria multiflora Hydro-alcoholic Extract on Bovine *Escherichia coli*. *Egypt. J. Vet. Sci.*, **53**(1), 139-145(2022).
5. Levinson, W. Review of medical microbiology and immunology. *McGraw-Hill Education*, (2014).
6. Abbas-Al-Khafaji, Z.K. and Aubais-aljelehawy, Q.H. Evaluation of antibiotic resistance and prevalence of multi-antibiotic resistant genes among *Acinetobacter baumannii* strains isolated from patients admitted to alyarmouk hospital. *Cellular, Molecular and Biomedical Reports.*, **1**(2), 60-68(2021).

7. Harris, M., Fasolino, T., Ivankovic, D., Davis, N.J. and Brownlee, N. Genetic Factors That Contribute to Antibiotic Resistance through Intrinsic and Acquired Bacterial Genes in Urinary Tract Infections. *Microorganisms*, **11**(6), 1407(2023).
8. Waffa, G.J. and Aqeel, M.Sh. Antibiotic Resistance Patterns of Escherichia coli Isolated From Broiler Chickens with Colibacillosis in Duhok Province. *Egypt. J. Vet. Sci.*, **54**(1),137-148 (2023).
9. Scheutz, F., Teel, L. D., Beutin, L., Piérard, D., Buvens, G., Karch, H. and Strockbine, N.A. Multicenter evaluation of a sequence-based protocol for subtyping Shiga toxins and standardizing Stx nomenclature. *Journal of Clinical Microbiology*, **50**(9), 2951-2963(2012).
10. Al Rekaby, S. M. The efficiency of enteric lactobacillus in preventing hemorrhagic colitis and blocking Shiga toxins productions in rats models infected with entero-hemorrhagic escherichia coli (ehc). *Iraqi Journal of Agricultural Sciences*, **52**(6), 1346-1355(2021).
11. Haque, M., Wang, B., Mvuyekure, A.L. and Chaves, B.D. Growth behavior of Shiga toxin-producing Escherichia coli, Salmonella, and generic E. coli in raw pork considering background microbiota at 10, 25, and 40° C. *International Journal of Food Microbiology*, **391**,110-134(2023).
12. Ali, R.M. and Saevan S.A. Effects of Probchick On E. ColiO157:H7 Experimental Infection in Broilers. *Egypt. J. Vet. Sci.*, **53**(3), 367-379 (2022).
13. Quinn, P.J., Markey, B.K., Carter, M.E., Donnelly, W.J. and Leonard, F.C. Veterinary Microbiology and Microbial Disease. *1st ed. Blackwell Science Ltd; London*. 163-167(2002).
14. Paton, A.W. and Paton, J.C. Detection and characterization of Shiga toxigenic Escherichia coli by using multiplex PCR assays for stx1, stx2, eaeA, enterohemorrhagic E. coli hlyA, rfbO111, and rfbO157. *J. Clin. Microbiol.*, **36**, 598-602(1998).
15. Taha, Z.M. and Yassin, N.A. Prevalence of diarrheagenic Escherichia coli in animal products in Duhok province, Iraq. *Iran. J. Vet. Res. Shiraz Univ.*, **20**(4),255-262(2019).
16. Alttai, N.A., Alsanjary, R.A. and Sheet, O.H. Genetic diversity of Escherichia coli harboring virulence gene Stx1 and Stx2 isolated from common carp fish in Nineveh Governorate using ERIC-PCR. *Iraqi Journal of Veterinary Sciences*, **37**(3), 701-705 (2023).
17. Gufe, C., Canaan, H.T., Mbonjani, B., Majonga, O., Marumure, J., Musari, S. and Machakwa, J. Antimicrobial Profiling of Bacteria Isolated from Fish Sold at Informal Market in Mufakose, Zimbabwe. *International Journal of Microbiology*, **2**,1-7 (2019).
18. Mahmmoud, E.N. and Aldabbagh, S.Y. Detection of extended spectrum beta lactam producing Escherichia coli isolated from Cyprinus carpio in Mosul city. *Iraqi Journal of Veterinary Sciences*, **36**,85-89(2022).
19. Kumar, H.S., Parvathi, A., Karunasagar, I. and Karunasagar, I. Prevalence and antibiotic resistance of Escherichia coli in tropical seafood. *World Journal of Microbiology and Biotechnology*, **21**(5), 619-623(2005).
20. Tyasningsih, W., Ramandinianto, S.C., Ansharieta, R., Witaningrum, A.M., Permatasari, D.A., Wardhana, D.K. and Ugbo, E.N. Prevalence and antibiotic resistance of Staphylococcus aureus and Escherichia coli isolated from raw milk in East Java, Indonesia. *Veterinary World*, **15**(8),2021-2028 (2022).
21. MS, S., Hassan, M.A., Hassnien, F.S., Abdel-Aal, M.M., Zakar, A.H. and Elshfey, S.A. Prevalence of Escherichia coli in fish obtained from retail fish markets in Gharbia Governorate, Egypt. *Benha Veterinary Medical Journal*, **34**(1), 254-260(2018).
22. pradhan, S., Pellino, C., Macmaster, K., Coyle, D. and Weiss, A.A. Shiga toxin mediated neurologic changes in murine model of disease. *Front. Cell. Infect. Microbiol.*, **6**,114(2016).
23. Manfredi, E., Rocca, M.F., Zintgraff, J., Irazu, L., Miliwebsky, E., Carbonari, C. and Chinen, I. Rapid and accurate detection of Shiga toxin-producing Escherichia coli (STEC) serotype O157: H7 by mass spectrometry directly from the isolate, using 10 potential biomarker peaks and machine learning predictive models. *Journal of Medical Microbiology*, **72**(5),001675(2023).
24. Gambushe, S.M., Zishiri, O.T. and El Zowalaty, M. E. Review of Escherichia Coli O157: H7 prevalence, pathogenicity, heavy metal and antimicrobial resistance, African perspective. *Infection and Drug Resistance*, 4645-4673 (2022).
25. Takashima, F. and Hibiya, T. An atlas of fish histology: normal and pathological 440 features, 2nd ed. *Tokyo, Kodansha*, (1995).
26. Suhaim, R.R. Fate of *Escherichia coli* O157: H7 on channel catfish (*Ictalurus 433 punctatus*) as affected by harvesting and processing schemes (Doctoral dissertation, uga) (2003).

## الكشف عن بكتيريا الإشريشيا القولونية المقاومة لطيف واسع من المضادات الحيوية والحاملة للجينين STX1 و STX2 من سمك الكارب الشائع (*Cyprinus carpio*) في محافظة صلاح الدين

قصي صالح جمعه

فرع الأمراض والدواجن - كلية الطب البيطري - جامعة تكريت - العراق.

تعتبر الإشريكية القولونية من أهم البكتيريا الملوثة للمزارع السمكية وتؤدي إلى تلوث وفساد الأسماك مما يشكل خطراً على الصحة العامة، هدفت الدراسة الحالية إلى إيجاد انتشار بكتيريا الإشريشيا القولونية المقاومة لطيف واسع من المضادات الحيوية التي تحمل STX1 و STX2 من أسماك الكارب الشائع (*Cyprinus carpio*). في محافظة صلاح الدين، ولهذا الغرض تم جمع 100 عينة من الأسماك، وتم استخدام الطرق التقليدية والوراثية. أظهرت نتائج الدراسة الحالية أنه من أصل 100 عينة من الأسماك تم عزل 41 عينة من الإشريكية القولونية بنسبة 41% و 19 عينة من أصل 48 عينة تم تشخيصها على أنها *ESBL E.coli* بنسبة 39.5% حسب اختبار PCR لجين Stx1. تم الكشف عن 31 عينة من أصل 48 عينة بنسبة 64.5% بينما تم الكشف عن جين Stx2 على 39 عينة بنسبة 81.2%.

الاستنتاجات: وجود نسبة تلوث عالية ببكتيريا الإشريشيا القولونية في الأسماك واحواضها في محافظة صلاح الدين، أغلب العزلات كانت مقاومة لطيف واسع من المضادات الحيوية. الجين STX2 أكثر ظهوراً من الجين STX1

**الكلمات الدالة:** الإشريكية القولونية، الشبوط الشائع، سموم شبيكا، المقاومة لطيف واسع من المضادات الحيوية