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# Paraovarian Cyst in Repeat-breeder Buffalo heifers: Ultrasonographic, Antioxidants and Serum Cell-free RNA markers

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THIS study aimed to (1) identify the status of reproductive tract of repeat-breeder heifers (RBH) ultrasonography, as well as (2) determine the variations in steroid hormones, oxidative stress biomarkers, and cell-free RNA in serum. These culled animals from 400 heifers at the breeding stationwere examined ultrasonography, and whole blood samples were collected. Theultrasonic diagnosed heifers as paraovarian cyst (POC, n=18) associated with genital pathological conditionswere compared with groups of control heifers at follicular(n=6), luteal (n=6), and early pregnant (n=10). The POC appeared as anechoic circumscribed masses located nearby, but separated from the ovary. Follicular number, corpus luteum size, serum cell-free RNA, and ovarian steroids were not affected by POC, although dominant follicle size tended to be lower (P=0.069)in POC withCL absence (POC-CL). Heifers with POC-CLhad lower MDA levels than pregnant ones. Buffaloes that had POC in the presence of CL (POC+CL) were found to have higher SOD activity than luteal and pregnant cases. Total serum proteinslevels recorded lower values n POC+CL compared to that in follicular phase. There was a significant positive relationship between POCsize, and GPx activity and CL size, but negative correlations with dominate follicles size. The POC might interfere with ovarian function, via alterations in antioxidant levels.

Keywords: Paraovarian cyst; Repeat-breeder buffalo heifers;Ultrasound;Antioxidant; Serum cell-free RNA

## **Introduction**

Knowledge about reproductive disorders in buffaloes is necessary due to its intimate relationship with animal reproductivity and productivity. Pathological conditions disrupting ovarian functions are the highest among the abnormalities of reproductive organs present in buffaloes.

A repeat breeder is generally defined as any cowthat has not conceived after three or more services associated with true estrus[1]. In buffaloes, the incidence of repeat breeding is low, although in different publications the incidence varied from 0.70% to 30%[2]. In field, both of anestrus as well as repeat breeding are the two major reasons of reproductive disorders in buffaloes [3].

The incidence of paraovarian cysts (POC) in buffaloes varies from 0.43-13.0% [4-9]. It wasstated that the incidence of POC was the highest recorded among ovarian affections in culled infertile buffaloes (13.74%), with the lowest incidence of luteal and follicular cysts (1.53 and 0.76%, respectively)[8]. Many

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authors assumed the low significance of POC in relation to buffalo fertility. Nevertheless, POC in other species like ewes causes infertility due to compress the uterine tube or close the opening of oviduct fimbria[10-11].

Peri-ovarian or para-ovarian cysts are fluidfilled structures in the vicinity of ovaries that varied in size from a few millimetres to that of a modest-sized follicle [12]. The cysts arise from mesothelial, mesonephric (Wolffian) or paramesonephric (Müllerian) structures contiguous normal ipsilateral ovary [13]. The POCare considered remnants of the mesonephric ducts. They are usually found at either the cranial or caudal pole of the ovary (epoophoron and paraoophoron cysts) and emerging from either the cranial or caudal sections of the mesonephric tubules[14].Furthermore, POC may sometimes be mistaken for cystic ovary on rectal palpation because of closeness to the ovary. The difference in detection of POC pre-slaughter per rectum and after slaughter reaches to 5% [8]. They are occasionally found around the ovary and fallopian tubes, attached in the broad ligaments of cows and buffaloes ([15]. The POC can be near the fimbria, alongside the uterine tubes, inside the broad ligament or mesosalpinx[10,16,17]. Tinyparaovarian cysts of few millimetre diameters are incidental findings in slaughtered buffaloes [18]. The cysts may vary in size from 1 to 5 cm in diameter and are usually round or oval in shape [9].

A proper balance between reactive oxygen species (ROS) that generate as by-products during different ovarian physiological process and antioxidants are essential for optimal ovarian functions during entire reproductive life in females [19].Oxidative stress is known as one of the most common mechanisms associated with natural events as pregnancy [20] and a variety of diseases [21]. It was shown that catalase (CAT), glutathione peroxidase (GPx), total antioxidant capacity (TAC), super oxide dismutase (SOD), and the lipid peroxide (malondialdehyde, MDA) play an important roles in ovarian functions (folliculogenesis, ovulation, and different luteal phases) [19].

Recently, serum cell-free RNA is known as key player for regulation of gene expression in different physiological conditions, and any alterations in their expression may be associated with pathological disorders [22]. Moreover, serum cell-free RNA has the potential to serve as

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biomarker for several medical conditions, because of their high stability [23].However, limitations in the isolation methods of serum cell-free RNA, as well as a lack of knowledge on factors, which may affect recovery rate of serum cell-free RNA from different biological samples, can hinder their successful discovery[24].Many studies reported that serum cell-free RNA is a potential biomarker for diagnosis of ovarian endometriosis [22, 25, 26].

Little is known about the changes inantioxidant parameters and serum cell-free RNA concentrations that may be associated with POC in buffalos. The hypothesis is the serum cell-free RNA is associated with the disease and POC is neither a major nor a secondary cause of infertility in buffaloes.We need to explain the association of POC withinfertilityin the reported buffalo heifers. Therefore, the current study aimed to (1) identify the status of reproductive tract of repeat-breeder heifersultrasonography, as well as (2) investigate the variations in ovarian structures, steroid hormones {estradiol (E2) & progesterone (P4)}, oxidative stress biomarkers(CAT, GPx, SOD, TAC, and MDA), and serum cell-free RNA, in these culled buffalo heifers.

## Materials and Methods

#### Animals

This study was carried out on eighteenPOCnon pregnantbuffalo heifers (Bubalusbubalis) aged more than three years. The heifers were excluded from breeding during routine examination by ultrasonography pregnancy diagnosis from 400 heifers. By repeating examination, they reported as non-pregnant and all were repeat breeders. These heifers belonged to a breeding station at MahaletMussa near to Sakha, Kafr El-Sheikh Governorate. All pregnant and non-pregnant heifers were subjected to the same management and nutrition programs. Bloodsamples were collected for measuring serum P4 and E2 hormones, TAC, MDA, GPx, SOD,CAT,total proteins and serum cell-freeRNA. Ethical considerations during blood samples were followed according to National Research Centre Committee.

### Ultrasound examination

Ultrasonographic examination was conducted per rectum in all buffaloes prior to blood sampling, using Sonoscape A6V supplied with 2-12 MHz linear transducer [27].To identify POC properly, ovarianboundaries and its parenchymaweredelineated, and POC appeared as an anechoic structure located alongside the ovary [28]. According to ovarianfindings by ultrasound; the animals were classified into: POC with no CL (corpus luteum)on the ovaries (POC-CL; n=10) and POC with CL on the ovaries (POC+CL; n=8), and were compared with groups of control heifers at follicular (n=6), luteal (n=6) and two months pregnancy (n=10). The association of POC with pathological conditions in the genital system (Vaginitis, cervicitis, endometritis, salpingitis, and oovaritis) is illustrated in Table (1).

## Measurement of ovariansteroidhormonesin serum

The concentrations of P4 and E2hormones were determined in serum, using commercial ELISA kits (ChemuxBioScience, South San Francisco, USA) as indicated by the manufacturer's instructions. The value of optical density (OD) was checked, using ELISA microplate reader (BioTek ELx800, Vermont, USA), at 450 nm wavelength for both steroidal hormones.

# Measurement of antioxidants and total protein in serum

The level of TAC was determined colorimetricallyvia commercial kit (Biodiagnostic, Egypt) as recorded by Koracevic et al. [29]. The MDA level was measured colorimetrically, using commercial kit (Biodiagnostic, Egypt) according to the method of Ohkawa et al.[30]. The activity of GPxandSOD was estimated kinetically, using commercial assay kits (Biodiagnostic, Egypt)according to Paglia and Valentine [31], and Nishikimi et al. [32], respectively. Total proteins were measured colorimetrically by Biuret method via a commercial kit (Biodiagnostic, Egypt) indicated by Gorna et al.[33].

## Serum cell-free RNA

For purification of cell-free total RNA, we used a miRNeasy serum/plasma kit (Qiagen). 200µl of thawed serum samples on ice were used for purification cell-free total RNA according to manufactures protocol. During purification steps, 3.5 µlof lyophilized C. elegans miR-39 miRNA mimic (miRNeasy Serum/PlasmaSpike-In Control) were added, at concentration  $1.6 \times 10^8$ copies/µl (working solution). In order to elute RNA,14µl of RNase-free water was added to the centre of the spin column membrane. Afterwards, the purified cell-free total RNA waskept at -80°C for further processing. The concentration of total RNA was checked by Nano-drop 2000/c (Thermo Fisher Scientific, Wilmington, USA), and also the RNA integrity was evaluated bydenaturing agarose gel electrophoresis and ethidium bromide staining.

## Statistical analysis

The obtained data were tabulated, presented as mean±Standard error of means (SE), and statistically analyzed with IBM SPSS version 21. Data were analyzed statistically using Mann-Whitney U test to define significance among two groups, while Kruse-William test was used to verify the significance between more than two groups. The normal distribution was checked via Shapiro-Wilk test and Gaussian distribution; and all data was passed normality test (alpha=0.05), (GraphPad Software, Inc., San Diego, CA, USA).Spearman correlation was used to define the correlation between the tested parameters. *P*-value was set at <0.05 to define statistical significance.

Numbers of animals with POC	Associated pathological conditions
1	Vaginitis
2	Mild cervicitis
3	Mild endometritis and oovaritis and salpingitis
1	Mildendometritis, ovarian tumour(22.7 x 18.8 mm)
2	Enlarged valva, cervicitis, follicular cyst (30.8 x 23.3 mm)
1	Pyosalpinix and uterine adhesions
3	Vaginitis, chronic cervicitis, pyosalpinex
2	Chronic fibrosis of cervix
3	Mild endometritis

 TABLE 1. Paraovariancyst (POC) buffaloes with description for the associated pathological conditions in the genital system.

#### <u>Results</u>

## Ultrasound findings

Representative ultrasound images of buffaloes had POC in presence of CL or absence is shown in (Fig. 1 and 2).Ovarian changes in animals demonstrated paraovarian cyst versus normal reproductive physiological statuses (cyclicity and pregnancy) is shown in (Table 2). Analysis of the obtained data revealed, as expected, that the paraovarian cysts size, follicular number (P= 0.845) and corpus luteum size (P= 0.161) were not statistically varied, although dominant follicle size tended to be significantly (P= 0.069) differed between groups.



Fig. 1. Representative ultrasonogram of paraovarian cyst in buffalo heifer nearby the ovary that presented corpus luteum. Paraovarian cyst (POC) appeared as a large translucent anechoic cavity. The corpus luteum (CL) showed a varying degree of heterogenicity.



Fig. 2. Representative ultrasonogram of paraovarian cyst in buffalo heifer nearby the ovary that presented growing follicles. Paraovarian cyst (POC) appeared as a large translucent anechoic cavity, and the growing follicle appeared smaller in size circumscribed anechoic areas within the ovarian parenchyma.

	Paraova	rian cyst	Normal <b>j</b>	physiological con	ditions	
	Without CL (POC-CL)	With CL (POC+CL)	Follicular	Luteal	Pregnant	<i>P</i> value
Paraovarian cyst size	49.23±6.01	30.50±5.49				0.10*
Dominant follicle size (mm)	7.2±0.96	14.90±2.31	16.5±1.32	8.71±2.51	7.95±0.26	0.069
Follicle number on ovaries	3.0±0.9	2.3±0.3	3.0±0.6	3.0±0.5	2.3±0.6	0.845
Size of CL (mm)		21.40±1.44		20.33±0.55	23.3±0.98	0.161

TABLE 2.	Ovarian	changes in	buffaloes	with	paraovariancyst.

\* Mann Whitney test

*Alterations in steroidal hormones, antioxidants, and serum cell-free RNA levels* 

Ovarian steroid hormone analysis (Table 3) showed significant (P= 0.003) difference in progesterone level between non-luteal (POC-CL and follicular phase) and luteal (POC+CL, cyclic CL, pregnancy). In contrary, estradiol variances between these groups did not reach statistical level (P= 0.160).

The level of antioxidants (Table 4) showed marked differences in MDA and SOD and total serum proteins (P < 0.009, 0.013, and 0.012, respectively), heifers having in paraovarian cyst compared to cyclic (follicular and luteal phases) and pregnant buffaloes. Cases with POC in the absence of CL (POC-CL) had lower MDA levels than pregnant cases (5.99±0.35 and 13.70±0.63 nmol/mL, respectively). Buffaloes had POC in the presence of CL (POC+CL) had higher SOD activity than luteal and pregnant cases (226.70±1.44U/ml vs. 166.90±3.47 and 168.20±1.29 U/ml, respectively). Total protein level was lower in POC+CL than those in follicular phase (3.69±0.39 and 8.39±0.31 g/dl, respectively).

Although serum free RNA not significantly varied between POC cases and physiological conditions (Fig. 3), it was significantly (P= 0.006) lower during pregnancy than follicular phase (7.58±0.34 vs. 14.92±0.44 ng/µl, respectively).

#### **Discussion**

Ovarian cysts are classified into intraovarian cysts, the cysts within the ovary, and periovarian (paraovarian) cyststhat are outside to the ovary. The POC are recorded in buffalo post-slaughter [34]. However, in our study;the POC was diagnosed in repeat-breeder buffalo heifers, by means of ultrasonography, and the incidence was 4.5%. Large POC may predispose for ovarian torsion and rupture of broad ligament. Concurrently, it was recorded that POC at an incidence of 4.4% of the examined slaughtered buffaloes[15]. Moreover, Vala[8] showed that the incidence of POC was the highest among ovarian affections in culled infertile buffaloes (13.74%), while luteal and follicular cysts incidences were the lowest (1.53 and 0.76%, respectively).

Former studies differed in the interpretation of the significance of POC on animal reproduction. While some of them [15] declared its nonsignificant effect on female fertility and its interference only with the diagnosis of ovarian cyst, others [10] showed that the fertility is distressed in case of the POC when compressing the uterus to block the oviductal opening. In this study most of the POC cases were encountered in animals culled due to subfertility problems.

Ultrasonographic measuring of POC showed that its size varied from 14.16 mm to 61.00 mm with mean values of  $49.23\pm6.01$  and  $30.50\pm5.49 \text{ mm}$  in case of CL presence or absence, respectively. This finding was in accordance with former studies showed that POC of a few millimetres in diameter are incidental findings in slaughtered buffaloes [18], though it may vary in size from 1 to 5 cm in diameter [9]. Bohîlţea et al. [36] indicated the use of ultrasound exam for paraovarian cyst diagnosis that appeared as a round or oval well-defined cystic mass, located close to the ipsilateral ovary, but clearly separated.

	P	araovarian cyst		Normal physiolog	gical conditions	
	Without CL (POC-CL)	With CL (POC+CL)	Follicular	Luteal	Pregnant	<i>P</i> value
P4 Conc. (ng/ml)	0.419±0.004 <sup>b</sup>	4.333±0.066 <sup>ab</sup>	0.293±0.049 <sup>b</sup>	2.597±0.264 <sup>ab</sup>	4.662±0.103ª	0.003
E2 Conc. (pg/ml)	2.668±0.063	4.077±0.921	5.440±0.385	3.080±0.246	3.212±0.496	0.160
P4/E2 ratio	0.155±0.005 <sup>ab</sup>	1.267±0.303 <sup>ab</sup>	0.053±0.012 <sup>b</sup>	$0.857{\pm}0.127^{ab}$	1.540±0.152ª	0.006

Kruse-William test.

Small different letters in the rows denote significantly (P<0.05) between groups

TABLE 4. Effect of paraovarian cyst on antioxidant parameters in buffaloes.
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	Paraov	arian cyst	Normal	physiological co	nditions	
	Without CL (POC-CL)	With CL (POC+CL)	Follicular	Luteal	Pregnant	<i>P</i> value
TAC (mM/L)	1.56±0.01	1.49±0.04	1.49±0.12	1.52±0.14	1.52±0.03	0.718
MDA (nmol/mL)	7.71±1.02 <sup>ab</sup>	5.99±0.35 <sup>b</sup>	9.61±0.59 <sup>ab</sup>	6.55±0.29 <sup>ab</sup>	13.70±0.63ª	0.009
GPx (mU/ml)	46.30±2.01	41.36±1.18	45.05±0.58	44.27±0.37	44.07±0.68	0.218
SOD (U/ml)	226.70±1.44ª	199.60±15.19 <sup>ab</sup>	173.30±4.41 <sup>ab</sup>	166.90±3.47 <sup>b</sup>	168.20±1.29 <sup>b</sup>	0.013
Total protein (g/dl)	3.69±0.39 <sup>b</sup>	5.74±1.28 <sup>ab</sup>	8.39±0.31ª	7.54±0.29 <sup>ab</sup>	5.09±0.51 <sup>ab</sup>	0.012

Kruse-William test

Small different letters in the rows denote significantly (P<0.05) between groups

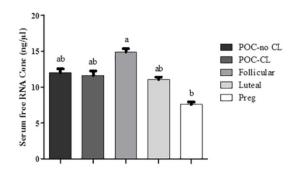


Fig. 3. The concentration of serum cell-free RNA in buffaloes demonstrated paraovarian cyst compared to control animals. Values (Mean±SE) with different letters were significantly different at *P*< 0.05 (Kruse-William test).

The interrelationship between examined parameters were analyzed and presented in (Table 5). There was a significant correlations between POC and GPx activity (r=0.586, P< 0.05) and CL size (r=0.817, P< 0.01), but negative correlation with dominate follicle size (r=-503, P< 0.01).

TAC         GP         Protein         Pd         Eq         Motion (minor)         Collicity         Collicity (minor)         Collicity (minor)         Street (minor)           MDA         0.166         -0.006         -0.443         0.078         0.311         0.058         -0.024         -0.026         -0.025         -0.033         0.235         -0.034         -0.026         -0.025         0.443         0.013         -0.036         -0.034         -0.026         -0.026         -0.026         -0.026         -0.035         0.235         -0.034         -0.253         -0.255         -0.035         -0.255         -0.035         -0.256         -0.035         -0.256         -0.035         -0.256         -0.035         -0.256         -0.035         -0.256         -0.035         -0.255	TABLE 5.	Spearman's	s correlation	s between a	ntioxidant pa	ırameters, s	teroid horn	nones, and	ovarian acti <sup>,</sup>	TABLE 5. Spearman's correlations between antioxidant parameters, steroid hormones, and ovarian activity in buffaloes.			
-0.253       0.226         -0.497       0.209         -0.454       -0.075         -0.301       0.084         -0.301       0.084         -0.267      879-**         -0.267       0.653         -0.227      879-**         -0.227      753-*         -0.212       .787*         -0.227      828-**         -0.295       -0.155         -0.295       -0.155		TAC	GPx	SOD	Protein	P4	E2	P4/E2 ratio	RNA	Dominant folliclesize (mm)	Follicle number on ovaries	CL size (mm)	Size of paraovarian cyst (mm)
-0.497 0.209 0.454 -0.075 -0.301 0.084 0.242879-** -0.267 0.653 -0.257877* 0.022753-* -0.212787* -0.295 -0.155 -0.295 -0.155	MDA	0.168	-0.006	-0.443	0.078	0.311	0.058	0.202	-0.352	-0.074	-0.253	0.226	-0.25
0.454 -0.075 -0.301 0.084 0.242879-** -0.267 0.653 0.022753-* -0.212 .787* 0.227828-** -0.295 -0.155 -0.487	TAC		0.022	0.435	-0.277	-0.045	-0.197	-0.065	-0.021	0.019	-0.497	0.209	0.296
-0.301 0.084 0.242879-** -0.267 0.653 0.022753-* -0.212 .787* 0.227828-** -0.155 -0.155	GP <sub>X</sub>			-0.007	0.121	-0.295	0.00	-0.368	0.085	-0.154	0.454	-0.075	.586*
0.242879-** -0.267 0.653 0.022753-* -0.212 .787* 0.227828-** -0.295 -0.155 -0.487	SOD				403-*	-0.338	-0.246	-0.324	0.424	0.141	-0.301	0.084	0.321
-0.267 0.653 0.022753-* -0.212 .787* 0.227828-** -0.295 -0.155 -0.487	Protein					-0.32	.725**	-0.352	0.382	0.464	0.242	879-**	-0.714
0.022753-* -0.212 .787* 0.227828-** -0.295 -0.155 -0.487	P4						-0.238	.916**	845-**	-0.222	-0.267	0.653	-0.335
-0.212 .787* 0.227828-** -0.295 -0.155 -0.487	E2							474-*	0.398	.694**	0.022	753-*	-0.739
0.227828-** -0.295 -0.155 -0.487	P4/E2								810-**	-0.311	-0.212	.787*	-0.148
-0.295 -0.155 -0.487	RNA									0.419	0.227	828-**	0.071
-0.487	DF size										-0.295	-0.155	503-*
	Fol. no.											-0.487	0.63
DF: Dominant Follicle. *;Correlation is significant at the 0.05 level (2-tailed). **;Correlation is significant at the 0.01 level (2-tailed).	CL size												0.817**
	DF: Dominar	tt Follicle. *;	Correlation i	s significant	at the 0.05 lev	vel (2-tailed)	. **;Correlat	tion is signif	ficant at the 0.	.01 level (2-tailed)			

We monitored for the first time some biochemical changes in association with its presence including lower MDA, total protein, and an increase in SOD. Antioxidants are responsible for the balance between reactive oxygen species production and clearance. The disturbance in this balance can persuade pathological ovarian dysfunction, which can ultimately influence fertility. Biochemical alternations associated with POC was first to be indicated herein where MDA and total proteins levels were lower than its corresponding values in pregnancy and follicular phase, respectively, while SOD activity was higher than luteal and pregnancy. Moreover, total serum protein levels were negatively correlated with SOD. POC size was positively correlated with GPx activity (r=0.586, P < 0.05) and negative correlated with dominate follicle size (r=-503, P <0.01). At the same time, total serum protein was positively correlated with E2. This might indicate that high SOD possibly will lower total protein and E2. The E2 is a product of follicles while it grow and responsible for the expression of the signs of heat. Former studies indicated that SOD had inhibitory impacts on estrogen synthesis by inhibiting FSH-induced aromatase activity in cultured granulosa cellsthat occurred at post-FSH receptor sites in rodentgranulosa cells in vitro[37]. Taken together, POC might predispose to ovarian dysfunction and low expression signs of heat mediated through oxidative stress. Although this relationships were not proven but our data might enthuse this assumption and explain why POC is largely recorded in culled animals, due to subfertility problems. It has been proposed that decreased antioxidant defence systems preludes to pathological consequences include many reproductive diseases such as cystic ovaries, endometriosis, and unexplained infertility [19].

In our study, the concentration of serum cellfree RNA was not significantly varied between POC cases and physiological conditions. This may indicate that POC has no a clear effect on the concentration of serum cell-free RNA, in the studied animals. Although, it was shown clear alterations at the expression profile of serum cell-free RNA in patients with some infertility problems such as; endometriosis and POC [25], this may indicate that POC might affect the expression profile of serum cell-free RNA, not its concentrations. Here in the current study, we did not go further for expression profile, due to limitation in our facility. We also noticed that the serum cell-free RNA level was significantly lower

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during pregnancy than follicular phase. This could be resulted from contribution of serum cell-free RNAin placental formation during pregnancy progression [38].

#### **Conclusions**

POC has mild or no effect on buffalo fertility as no clear changes was found in steroid levels as well as serum cell-free RNA concentration. However, we observed alterations in some antioxidant parameters, which might interfere with ovarian functions in the examined buffalo cases.

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التكيس خارج المبيض في عجلات الجاموس المتكررة الشياع: الموجات فوق الصوتية ، مضادات الأكسدة وعلامات الحمض النووي الريبوزي في المصل الخالي من الخلايا

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هدفت هذه الدراسة إلى (١) تحديد حالة الجهاز التناسلي لعجلات الجاموس المتكررة الشياع بالموجات فوق الصوتية ، وكذلك (٢) تحديد الاختلافات في هرمونات الستيرويد ، والمؤشرات الحيوية للتأكسد ، والحمض النووي الريبوزى في المصل الخالى من الخلايا. تم فحص الحيوانات المستبعدة من ٤٠٠ بقرة في محطة تربية العجلات بالموجات فوق الصوتية ، وحمع عينات دم. تم تشخيص العجلات بالموجات فوق الصوتية على أنها العجلات بالموجات فوق الصوتية ، وجمع عينات دم. تم تشخيص العجلات بالموجات فوق الصوتية على أنها العجلات بالموجات فوق الصوتية ، وكن على من الخلايا. تم فحص الحيوانات المستبعدة من ٤٠٠ بقرة في محطة تربية تكيس خارج المبيض (ن = ١٨) و كانت مرتبطة بحالات مرضية تناسلية وتم مقارنتها مع مجموعات ضابطة في الطور الجريبي (ن = ١) ، الأصفري (ن = ١) ، والحمل المبكر (ن = ١٠). ظهرت التكيسات كمتل محدده عيمة الصدى تقع بالقرب من المبيض ولكنها منفصلة عنه. لم يتأثر عدد الجريبات وحجم الجسم الأصفر والحمض النووي الريبوزى المعزول من المصل والستيرويدات المبيضية بوجودالتكيسات خارج المبيض لوالخر من المحل والستيرويدات المبيضية بوجود التكيسات خارج المبيض ولكنها منفصلة عنه. لم يتأثر عدد الجريبات وحجم الجسم الأصفر والحمض الرغم من أن حجم الجريب الى ألى يكون أقل (60.09) في غياب المبيض وحجم الجسم الأصفر كان لديها مستويات مال الحالات والحمض النووي الريبوزى المعزول من المصل والستيرويدات المبيضية بوجودالتكيسات خارج المبيض على موالم من أن حجم الجريبة السائده يميل إلى أن يكون أقل (60.09) في غياب الجسم الأصفر كان الحالات والحمض الرغم من أن حجم الجريبة السائده يميل إلى أن يكون أقل (60.09) في غياب الجسم الأصفر مان الحالات الرغم من أن حجم الجريبة السائده يميل إلى أن يكون أقل (60.09) في غياب الجسم الأصفر ما ألومل الحربيات فرجما الحوال الحمل الحمل وألفي العجلات ذات المربيات خارج المبيض ما الحول المصوبة في عليها مستويات خارج المبيض مع كان الحمل وأل في العجلات ذات التكيسات خارج المبيض مع وجود الجسم الأصفر والحوامل. كان تركيز البروتينات الكلية في المصل أقل في العجلات ذات التكيسات خارج المبيض مع وجود الجسم الأصفر والحوامل. كان تركيز البروتينات الكلية في المصل أقل في العجلات ذات التكيسات خارج المبيض ما وربود ها الحسان مناي علائة ماي مالحم الحريي الرصفرم والحل مع وطيفة الحريل مع ماع وربود اليم

الكلمات الدالة: التكيس خارج المبيض. عجلات الجاموس المتكررة الشياع ؛ الموجات فوق الصوتية. مضادات الأكسدة. الحمض النووى الريبوزى في المصل